

Direct Potentiometric Method for Human Stress Determination

Sakač, N.; Karnaš, M.; Grčić, M.

Source / Izvornik: **Chemical and Biochemical Engineering Quarterly, 2015, 29, 315 - 321**

Journal article, Published version

Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

<https://doi.org/10.15255/CABEQ.2015.2175>

Permanent link / Trajna poveznica: <https://urn.nsk.hr/urn:nbn:hr:151:616712>

Rights / Prava: [In copyright](#) / [Zaštićeno autorskim pravom.](#)

Download date / Datum preuzimanja: **2024-11-23**



Sveučilište Josipa Jurja
Strossmayera u Osijeku

**Fakultet
agrobiotehničkih
znanosti Osijek**

Repository / Repozitorij:

[Repository of the Faculty of Agrobiotechnical
Sciences Osijek - Repository of the Faculty of
Agrobiotechnical Sciences Osijek](#)



Direct Potentiometric Method for Human Stress Determination

N. Sakač*, M. Karnaš, and M. Grčić

Department of Chemistry, Josip Juraj Strossmayer University of Osijek, Cara Hadrijana 8A, 31000 Osijek, Croatia

doi: 10.15255/CABEQ.2015.2175

Original scientific paper
Received: September 9, 2015
Accepted: January 15, 2015

The application of platinum redox electrode for potentiometric determination of salivary amylase activity as a stress biomarker is described. The candidates were divided into two groups, physical activity – medium intensity exercise (ten candidates), and psychical activity – student exam (thirteen candidates). The experimental data were compared with an adapted theoretical model where the sensor and analyte properties were optimized using Solver and the least-squares criterion to fit a theoretical curve into the experimental data set. It was found that, in both groups, the salivary amylase activity had increased after exposure to stress activities. For physical stress, the salivary amylase activity increase was found to be up to 699 %, with a potential difference between the trained and “untrained” candidates’ dependence; and for psychical stress, the salivary amylase activity increase was dispersed in a range from 117 % to 1201 %. Proposed methodology offers a fast and inexpensive way to determine salivary amylase activity and stress levels in humans.

Key words:

stress, saliva, amylase, direct potentiometry

Introduction

Stress was firstly defined in 1935¹, as a reaction of humans to external stimuli. In the modern way of life and every day actions, the term stress is very common and has a significant influence on human reactions, health, work ability and mood. There is also a direct connection between stress and depression in humans². Apart from adults, recent studies show that preschool children are also under stress, and some helping support strategies could reduce it³. Thus, it appears that quantification of stress is a challenging issue, not only in humans, but also in animals⁴. Stress response is inducing the sympathetic nervous system and hypothalamic-pituitary-adrenal system, where hormone regulation is included (norepinephrine and cortisol). Invasive techniques measure hormone levels in the blood as an indicator of stress, but these levels are very low and hard to measure, and response time is too long and delayed by 20–30 minutes in response to loading stress.

The use of salivary biomarkers has gained increased popularity over the past decade in psychological and biomedical research. While the measurement of free cortisol in saliva has proven to be in correlation with stress⁵, salivary amylase activity has appeared as a new highlight in stress measurement⁶.

When secretion of the salivary amylase is stimulated by direct innervations, its response is very quick, generally within one to a few minutes, a markedly quicker response than that induced by hormonal regulation^{7,8}. Salivary glands amplify the low levels of norepinephrine and give a faster and more sensitive response to the psychological stress than cortisol. To date, there are no international standard measurement methods for human salivary amylase activity. Salivary amylase has a high activity, ranging from several thousands to hundred thousand units per litre⁹.

According to literature, in human and animal science, amylase activity rises remarkably under autonomic system activation¹⁰, such as response to a) physical stressors like exercise^{11,12}, exposure to a high-pressure chamber¹³, running¹⁴, bicycle exercise¹⁵, sky diving or cold exposure¹⁶; and b) psychological stress⁷ or relaxation interventions¹⁷. While it seems clear that alpha-amylase levels rise following physical stress, the response to psychological stressors appears to be more inconsistent. This might be due to the psychological nature of the stressors employed or other methodological details⁵. Furthermore, concerning medical aspects, salivary amylase activity has also been employed as a marker for schizophrenia, since several researches indicate that the autonomic nervous system dysfunctions in patients with schizophrenia¹⁸; in patients with chronic relapsing pancreatitis, the activity of the salivary amylase was significantly lowered¹⁹ and recently, there have been attempts to suggest salivary amy-

*Corresponding author: e-mail: nikola.sakac@gmail.com; phone: +38531399970

lase as a new biomarker for cardiovascular diseases, like heart failure.²⁰

Saliva collection methods have many variations and include absorbent materials, like cotton²¹; commercial serviettes; passive drooling and spitting, and other methods²².

Published methods for salivary amylase activity include variations of biosensors, usually glucose biosensors, like disposable screen-printed amylase biosensor based on ferrocene as an electron transfer mediator²³, a disposable point-of-care, colorimetric biosensor strip²⁴, biosensor microfluidic SIA systems²⁵, and spectrophotometric determination of salivary amylase activity with p-nitrophenyl maltoside as substrate²⁶. In addition, many other analytical methods for alpha amylase determination could be employed^{27,28,29}.

The aim of this investigation was to test a new methodology for direct potentiometric determination of salivary amylase, based on our previous work³⁰, and quantify human stress as a function of salivary amylase activity at different stress conditions: physical activity – exercise, and psychical (mental) activity – student exam.

Experimental part

Reagents

α -Amylase (EC 3.2.1.1) from *Aspergillus oryzae* (its activity was 36 U mg⁻¹) was purchased from Fluka (Switzerland) and used as the standard amylase reagent without purification. Soluble starch, calcium chloride, sodium chloride and iodine were purchased from Kemika (Croatia). Potassium iodide was purchased from Sigma-Aldrich (SAD), glacial acetic acid from Panreac (Spain), sodium acetate trihydrate (CH₃COONa · 3H₂O) from J. T. Baker (Holland). All chemicals were p.a. grade.

Solutions

Starch solution

An amount of 5 g of dried soluble starch was mixed in 0.1 mol L⁻¹ acetate buffer solution (pH 6.0) in 100 mL flask. After heating and stirring, the starch had completely dispersed. The solution was cooled to room temperature to perform the measurements. Starch solutions were prepared daily to avoid microbial degradation and retrogradation.

Conditioning solution

The conditioning solution was prepared in 500 mL volumetric flask by dissolving CaCl₂ (6 mmol L⁻¹) and NaCl (20 mmol L⁻¹) in a 0.1 mol L⁻¹ acetate buffer solution (pH 6.0).

Saliva sample solutions

Saliva sample solutions (1 %) were prepared by adding 0.5 mL of each saliva sample to a 50 mL volumetric flask and then diluting it to the mark with a conditioning solution. The resulting solution was stirred without heating until the saliva had completely dissolved. The conditioning solution was used to maintain the constant volume and stability. The addition of CaCl₂ and NaCl to the saliva solution serves to stabilise enzyme activity.

Standard α -amylase solution

A standard α -amylase solution was prepared and standardised as described by Sakac *et al.*³ An amount of 5 mg of α -amylase (Fluka, Switzerland (0.36 U mL⁻¹)) was dissolved in 500 mL of the conditioning solution.

Acetic acid-triiodide solution

The acetic acid-triiodide solution (ATIS) was prepared by dissolving solid iodine ($c = 100 \mu\text{mol L}^{-1}$) in potassium iodide solution ($c = 0.05 \text{ mol L}^{-1}$) and adding glacial acetic acid to the final concentration 1 mol L⁻¹. ATIS was used for the inhibition of ptyalin activity and the reaction of triiodide with the nonhydrolyzed starch.

Apparatus

A Metrohm 780 pH meter, a 728 Stirrer, a Metrohm 765 Dosimat (all from Metrohm, Switzerland), homemade software and a platinum redox electrode IJ64 (Ionode, Australia) were all used for the response measurements. A silver/silver (I) chloride reference electrode (Metrohm, Switzerland) with 3 mol L⁻¹ KCl inner solution was used as a reference. A thermostat (PolyScience, USA) was used for the amylase incubation.

Direct potentiometric measurements were performed on an eDAQ 186 Quad Amp pH/mV amplifier connected to an eDAQ ecorder 821 8-channel data acquisition system operated by the eDAQ Echem 1.5 software (all from eDAQ, Australia).

Groups and sampling

The experiment included two groups of people. The first group were candidates involved in physical activity – 30 minutes active exercise for young healthy adults, males and females, age range from 22–30 years. The saliva samples were taken before (relaxed) and immediately after training (stressed). The second group of candidates were focused on mental stress and included students in the time of writing an exam. Here a group of young healthy adults, age range from 20–24 years, were sampled

just before writing an exam (stress) and on a day at the same time when they had no college obligations (relaxed). We considered both the male and female population.

Saliva was sampled by cotton adsorbent attached on the capillary tube, placed under the tongue, and collected in a tube³⁰. Sampling took up to five minutes, and the result is transparent and filtrated saliva. The collecting tubes were stored in the refrigerator at $-20\text{ }^{\circ}\text{C}$ until analysis. The time lag between the occurrences of mentally stimulated salivary amylase was one to three minutes; the recovery of normal salivary amylase levels after stress reduction is very rapid: the duration of amylase elevation is only about 10 minutes²⁸, which makes this sampling method suitable.

Measurement of the salivary amylase activity

Test tubes containing the same volume of starch solution were heated in a water bath at controlled temperature of $45\text{ }^{\circ}\text{C}$. The buffered saliva solution was incrementally added to the starch solutions in test tubes at volumes ranging from 0 to 2.5 mL. The volume of the reaction mixture was held constant by a periodic addition of conditioning solution. After 10 minutes of heating at $45\text{ }^{\circ}\text{C}$, the reaction was terminated by adding 5 mL of ATIS. The incubation time was 10 minutes. Test tube solutions were cooled to room temperature using a water bath. Redox sensor was used for direct potentiometric measurement of the redox potential of the solutions. As a comparator, a standard solution of α -amylase was used, following the same procedure.

Data elaboration and optimization strategy using Solver

The experimental data were elaborated using MS Excel and a Solver add in. The calculations were based on the model previously presented³⁰. *Solver* is a spreadsheet optimization modelling system incorporated into Microsoft Excel that can be used for solving different linear and nonlinear problems.

Solver was activated by choosing *Add ins...* in the *Tools* menu. It was used to compare an array of data predicted by the model with an initial set of parameter values over a range of dependent variable values with a set of experimental data. Then, the sum of squared residuals (SSR) between the two arrays was calculated by varying the parameter values to minimize the error (SSR) between the two data sets.

The *Solver* optimization consists of the following steps:

a) Generation of a worksheet containing the data, fit with an independent variable E (redox potential in mV) and dependent variable $mAmy$ (mass of ptyalin in μg).

b) Insertion of a column containing E_{calc} , which was calculated by means of Eq. 7 to describe the response of the platinum amylase sensor and include the appropriate number of parameters to be varied (Changing Cells). These parameters include the sensor slope (S), the constant potential term ($Const$), and the proportionality factor (k). Instead of a very low k value (on the order $1 \cdot 10^{-4}$), the $\log k$ value should be used. Providing different sets of initial conditions ensured that *Solver* found a global minimum.

c) Insertion of a column for calculating the squares of the residuals, $E - E_{calc}$, for each data point.

d) Calculation of the sum of the squares of the residuals (Target Cell).

e) The use of *Solver* to minimize the sum of the squares of the residuals (Target Cell) by changing the selected parameters of Eq. 2 (Changing Cell). No constraints were applied to the variables.

The macro SolvStat provided the regression statistics for *Solver* by calculation of the standard deviations of the parameters, correlation coefficients and standard errors of the y estimate $SE(y)$.

Results and discussion

Modelling the response mechanism

Salivary amylase catalyses the hydrolysis of starch from a starch-triiodide complex and releases the triiodide ion. The increase in the triiodide ion concentration increases the triiodide/iodide redox couple ratio, resulting with an increase in the electrode potential of the redox sensor according to the Nernst equation³⁰.

$$E = E^0 + \frac{RT}{2F} \ln \frac{(C_{I_3^-})}{(C_{I^-})^3} = E^0 + S \log \frac{(C_{I_3^-})}{(C_{I^-})^3} \quad (1)$$

E^0 = constant potential term, S = slope of the sensor (mV/decade of activity), $(C_{I_3^-})$ = triiodide concentration, (C_{I^-}) = iodide concentration (considered to be constant).

Thus, the increase in the electrode potential relates to the ptyalin concentration/activity. The referred model has been adopted for salivary amylase activity determination.

Equation 2 represents the Nernst equation after salivary amylase addition to the system:

$$E = Const + S \log \left(C_{I_3^-} \right)_f \quad (2)$$

where $(C_{I_3^-})_f$ represents free triiodide concentration (mol L^{-1}), which is a summary of initial triiodide ions in solution $(C_{I_3^-})_0$ and a triiodide ion released by the salivary amylase activity $(C_{I_3^-})_l$.

This can be described:

$$(C_{I_3^-})_f = (C_{I_3^-})_0 + (C_{I_3^-})_l \quad (3)$$

Salivary amylase hydrolyses the starch and releases triiodide, which increase in concentration is proportional to the amylase quantity added:

$$(C_{I_3^-})_l = k \cdot m_{Amy} \quad (4)$$

where

k = proportionality factor

m_{Amy} = amylase quantity (μg).

By inserting Eq. (4) into (3), the following expression was obtained:

$$(C_{I_3^-})_f = (C_{I_3^-})_0 + k \cdot m_{Amy} \quad (5)$$

The salivary amylase quantity can be calculated from Eq. (5):

$$m_{Amy} = \frac{(C_{I_3^-})_f - (C_{I_3^-})_0}{k} \quad (6)$$

The adopted model was used for experimental data evaluation and salivary amylase activity calculation. A section of the spreadsheet that displays the po-

tentiometric measurements and the model parameters for the response characteristics of a platinum redox electrode sensor to exposure to the range of volumes of 1 % saliva solution is shown in Table 1. In Table 1, the Solver modelling using the proposed model and experimental data set is presented as a MS Excel spreadsheet. The calculated variables were slope (S), proportionality factor (k) and initial triiodide ion in solution $(C_{I_3^-})_0$. The Solver uses the least-squares criterion to fit a theoretical curve into the experimental data set, and the concentration of free triiodide, a triiodide ion $(C_{I_3^-})_f$ and released triiodide, a triiodide ion $(C_{I_3^-})_l$ could be calculated (Eq. (3)). The concentration of $(C_{I_3^-})_l$ at the beginning of the measurement was 0, since there was no amylase action. When the salivary amylase solution was added, the amylase took action, resulting in an increase in $(C_{I_3^-})_l$ and $(C_{I_3^-})_f$, respectively. Then, by using Eq. (6), the salivary amylase quantity and concentration could be calculated.

Following the model and Eq. 6, information on the amount of salivary amylase could be expressed. This is shown in Figure 1, where the logarithm of the amount of salivary ($\ln(m_A)$) amylase is dependent on the redox signal potential change. The linear equation for the corresponding dependence was $y = 0.1062x - 0.2492$, with the correlation coefficient (R^2) value 0.9828.

Table 1 – Selected potentiometric measurements and model parameters for the response characteristics of a platinum redox electrode sensor to exposure to a range of volumes of 1 % saliva solution. (*ssr* – sum of squared residuals; *sr* – squared residuals for the experimental data, *S* – sensor slope; E_0 – standard redox potential; E – experimental redox potential; $(C_{I_3^-})_f$ – free triiodide concentration (Eq. 5), C_{Amy} – salivary amylase concentration, $(C_{I_3^-})_l$ – liberated triiodide ion.

model parameters				C_{Amy}		
S	53.80	ssr	11.34	$\mu\text{g mL}^{-1}$	Units	
k	2.35E-05			17.76	0.639	
$C(I_3^-)_0$	2.26E-05					
$V(\text{saliva})/\text{mL}$	Experimental E/mV	$\Delta E = E - E_0$	$C(I_3^-)_f$	$C(I_3^-)_l$	Model E/mV	sr
0	289.9	0	1.00E-04	0	291.14	1.53
0.1	294.8	4.9	1.45E-04	4.48E-05	296.06	1.60
0.15	297	7.1	1.71E-04	7.09E-05	298.19	1.41
0.2	299.3	9.4	2.03E-04	1.03E-04	300.13	0.69
0.25	302.1	12.2	2.51E-04	1.51E-04	301.93	0.03
0.3	303.7	13.8	2.84E-04	1.84E-04	303.59	0.01
0.4	307	17.1	3.64E-04	2.64E-04	306.61	0.15
0.6	312.9	23	5.68E-04	4.68E-04	311.67	1.50
0.9	319.2	29.3	9.14E-04	8.14E-04	317.67	2.35
1.2	322.8	32.9	1.20E-03	1.10E-03	322.43	0.14
1.5	325	35.1	1.42E-03	1.32E-03	326.39	1.93

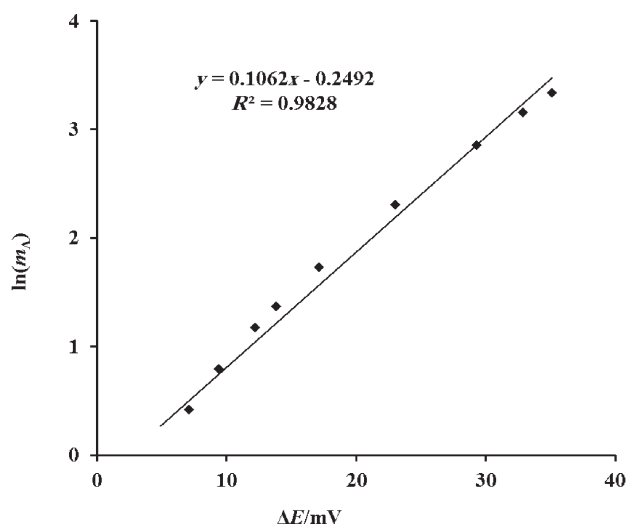


Fig. 1 – Logarithm of the amount of salivary ($\ln(m_A)$) amylase as a function of the redox signal potential change, according to model and Eq. (6)

Influence of physical activity

The influence of physical activity on the salivary amylase concentration was exemplified through the intensive exercise. Exercise intensity was medium. The investigation was carried out on a group of ten healthy young people.

Saliva was collected before the exposure to the intensive exercise, and after 30 minutes of training it was collected again. The salivary amylase concentration was measured before and after training using the previously presented model. The results of salivary amylase concentration before and after exercise are shown in Table 2.

The salivary amylase activity ranges from 4.51 to 34.04 $\mu\text{g mL}^{-1}$ for samples taken before training,

Table 2 – Results of salivary amylase concentration in saliva before and after exercise

Series	Before exercise (baseline)		After exercise (stress)		Increase (%)
	$\mu\text{g mL}^{-1}$	units	$\mu\text{g mL}^{-1}$	units	
1	4.51	0.162	8.89	0.320	197
2	12.22	0.440	20.79	0.749	170
3	12.99	0.468	29.96	1.079	231
4	27.45	0.988	62.84	2.262	229
5	22.88	0.824	91.53	3.295	400
6	11.70	0.421	48.95	1.762	418
7	9.35	0.337	65.36	2.353	699
8	34.04	1.225	51.25	1.845	151
9	11.31	0.407	23.78	0.856	210
10	25.22	0.908	43.00	1.548	170

and 8.89 to 91.53 $\mu\text{g mL}^{-1}$ for samples taken after training. The minimum increase in salivary amylase activity was shown in sample 8 (151 %) and the highest increase in sample 7 (699 %). Candidates 1, 2, 3, 4, 9, 10, 8 were trained regularly and their salivary amylase activity increase was quite lower than that of untrained candidates. According to the literature, there is no data available on the response difference in “basal” salivary amylase activity and “stress” salivary amylase activity between trained and untrained persons¹⁰. There is a good prospective to continue the research on that issue.

Influence of psychical (mental) activity

The influence of psychical (mental) activity on the salivary amylase concentration was exemplified through the intensive mental effort (student exam). The investigation was carried out on a group of thirteen healthy young people.

The salivary amylase activity ranges from 1.73 to 36.56 $\mu\text{g mL}^{-1}$ for samples taken in relax mode (baseline), and 11.79 to 64.92 $\mu\text{g mL}^{-1}$ for samples taken before the exam. The minimum increase in salivary amylase activity was shown in candidates 6 and 13 (117 % and 119 %) and the highest increase in candidate 8 (1201 %). Unfortunately, the final exam score was not calculated in the final amylase activity results shown in Table 3.

Table 2 confirms the literature results that salivary amylase levels rise following physical stress. According to literature⁵, the response to a psycho-

Table 3 – Results of salivary amylase concentration in saliva before student exam and in relaxation mode (no exam)

Series	No exam (baseline)		Before exam (stress)		Increase (%)
	$\mu\text{g mL}^{-1}$	units	$\mu\text{g mL}^{-1}$	units	
1	18.43	0.664	64.92	2.337	352
2	12.52	0.451	17.14	0.617	137
3	3.23	0.116	11.79	0.424	365
4	1229	0.442	33.84	1.218	275
5	1930	0.695	55.09	1.983	285
6	30.38	1.094	35.68	1.284	117
7	31.63	1.139	40.23	1.448	127
8	2.49	0.089	29.86	1.075	1201
9	36.56	1.316	43.62	1.570	119
10	5.46	0.196	26.96	0.970	494
11	25.35	0.912	44.98	1.619	177
12	1.73	0.062	13.87	0.499	804
13	25.22	0.908	29.99	1.080	119

logical (mental) stressor appears to be more inconsistent, and compared to physical activity, shows higher dispersion in salivary amylase activity increase (Table 3). It is not so simple to define the reason; it is a matter for discussion and further investigation with controlled experimental design.

Conclusion

Salivary amylase activity was determined using direct potentiometric method with redox sensor and starch-triiodide complex; and corresponding adapted model in real samples (candidates). Corresponding salivary amylase activity is in relation to human stress and was confirmed by the results for two groups of experiments: a) Physical stress (medium intensity exercise) resulted in relatively high salivary amylase activity increase (up to 699 %) with potential “trained level” dependence; and b) Psychical (mental) stress with dispersed salivary amylase activity increase values (ranging from 117 to 1201 %), which makes it more inconsistent and requires more objective and well-defined experimental groups and environment.

The proposed methodology offers a fast and inexpensive way to determine salivary amylase activity and stress levels in humans. The method has potential in checking body trained levels in sports.

References

1. Selye, H., McKeown, T., Studies on the physiology of the mental placenta in the rat, *Proc. Roy. Soc.* **B 119** (1935) 1. doi: <http://dx.doi.org/10.1098/rspb.1935.0073>
2. Hammen, C., Stress and depression, *Annual Review of Clinical Psychology* **1** (2005) 293. doi: <http://dx.doi.org/10.1146/annurev.clinpsy.1.102803.143938>
3. Hatfield, B. E., Hestenes, L. L., Kintner-Duffy, V. L., O'Brien, M., Classroom emotional support predicts differences in preschool children's cortisol and alpha-amylase levels, *Early Child. Res. Q.* **28** (2013) 347. doi: <http://dx.doi.org/10.1016/j.ecresq.2012.08.001>
4. Behringer, V., Deschner, T., Möstl, E., Selzer, D., Hohmann, G., Stress affects salivary alpha-amylase activity in bonobos, *Physiol. Behav.* **105** (2012) 476. doi: <http://dx.doi.org/10.1016/j.physbeh.2011.09.005>
5. Kirschbaum, C., Hellhammer, D. H., Salivary cortisol in psychoneuroendocrine research: Recent developments and applications, *Psychoneuroendocrinology* **19** (1994) 313. doi: [http://dx.doi.org/10.1016/0306-4530\(94\)90013-2](http://dx.doi.org/10.1016/0306-4530(94)90013-2)
6. Garrett, J. R., Effects of autonomic nerve stimulation on salivary parenchyma and protein secretion, in *Garrett, J. R., Ekstrfm, J., Anderson, L. C.* (Eds.), *Neural Mechanisms of Salivary Gland Secretion*, *Front. Oral Biol.* **11** (1999) 59.
7. Skosnik, P. D., Chatterton Jr., R. T., Swisher, T., Park, S., Modulation of attentional inhibition by norepinephrine and cortisol after psychological stress, *Int. J. Psychophysiol.* **36** (2000) 59. doi: [http://dx.doi.org/10.1016/S0167-8760\(99\)00100-2](http://dx.doi.org/10.1016/S0167-8760(99)00100-2)
8. Yamaguchi, M., Kanemori, T., Kanemaru, M., Takai, N., Mizuno, Y., Yoshida, H., Performance evaluation of salivary amylase activity monitor, *Biosens. Bioelectron.* **20** (2004) 491. doi: <http://dx.doi.org/10.1016/j.bios.2004.02.012>
9. Read, G. F., Tenovuo, J. O. (Eds.), *Human Saliva: Clinical Chemistry and Microbiology II*, Vol. 1, CRC Press Inc., Boca Raton, Florida., 1989, pp 1-54.
10. Rohleder, N., Nater, U. M., Determinants of salivary a-amylase in humans and methodological considerations, *Psychoneuroendocrinology* **34** (2009) 469. doi: <http://dx.doi.org/10.1016/j.psyneuen.2008.12.004>
11. Ehlert, U., Erni, K., Hebisch, G., Nater, U., Salivary alpha-amylase levels after yohimbine challenge in healthy men, *J. Clin. Endocrinol. Metab.* **91** (2006) 5130. doi: <http://dx.doi.org/10.1210/jc.2006-0461>
12. Chicharro, J. L., Lopez-Mojares, L. M., Lucia, A. Perez, M., Alvarez, J., Labanda, P., Calvo, F., Vaquero A. F., Overtraining parameters in special military units, *Aviat Space Environ Med.* **69** (1998) 562.
13. Gilman, S. C., Fischer, G. J., Biersner, R. J., Thornton, R. D., Miller, D. A., Human parotid alpha-amylase secretion as a function of chronic hyperbaric exposure, *Undersea Biomed. Res.* **6** (1979) 303.
14. Steerenberg, P. A., Asperen, I. A., Amerongen, A. N., Biewenga, J., Mol, D., Medema, G., Salivary levels of immunoglobulin A in triathletes, *Eur. J. Oral Sci.* **105** (1997) 305. doi: <http://dx.doi.org/10.1111/j.1600-0722.1997.tb00245.x>
15. Walsh, N. P., Blannin, A. K., Clark, A. M., Cook, L., Robson, P. J., Gleeson, M., The effects of high-intensity intermittent exercise on saliva IgA, total protein and alpha-amylase, *J. Sports Sci.* **17** (1999) 129. doi: <http://dx.doi.org/10.1080/026404199366226>
16. Chatterton Jr., R.T., Vogelsong, K. M., Lu, Y. C., Ellman, A. B., Hudgens, G. A., Salivary alpha-amylase as a measure of endogenous adrenergic activity, *Clin. Physiol.* **16** (1996) 433. doi: <http://dx.doi.org/10.1111/j.1475-097X.1996.tb00731.x>
17. Yamaguchi, M., Deguchi, M., Miyazaki, Y., The effects of exercise in forest and urban environments on sympathetic nervous activity of normal young adults, *J. Int. Med. Res.* **34** (2006) 152. doi: <http://dx.doi.org/10.1177/147323000603400204>
18. Ieda, M., Miyaoka, T., Wake, R., Liaury, K., Tsuchie, K., Fukushima, M., Araki, T., Ezoe, S., Inagaki, T., Horiguchi, J., Evaluation of autonomic nervous system by salivary alpha-amylase level and heart rate variability in patients with schizophrenia, *Eur. Arch. Psychiatry Clin. Neurosci.* **264** (2014) 83. doi: <http://dx.doi.org/10.1007/s00406-013-0411-6>
19. Grimm, K., Rossbach, G., Kasper, H., Amylase activity of parotid saliva in acute and chronic pancreatitis, *Acta Hepato-gastro.* **23** (1976) 333.
20. Suska, A., Alehagen, U., Lundstrom, I., Dahlstrom, U., Salivary Alpha-Amylase Activity, a New Biomarker in Heart Failure?, *J. Clin. Exp. Cardiol.* **2** (2012). doi: <http://dx.doi.org/10.4172/2155-9880.s2-005>
21. Harmon, A. G., Hibbel, L. C., Rumyantseva, O., Granger, D. A., Measuring salivary cortisol in studies of child development: Watch out-what goes in may not come out of saliva collection devices, *Dev. Psychobiol.* **49** (2007) 495. doi: <http://dx.doi.org/10.1002/dev.20231>
22. Navazesh, M., Methods for Collecting Saliva, *Ann. N.Y. Acad. Sci.* **694** (1993) 72. doi: <http://dx.doi.org/10.1111/j.1749-6632.1993.tb18343.x>

23. *Chao-Shi, Z., Mi, Z., Guo-Ming, X., Peng, L., Xing-Liang, X., Hua-Jian, X., Jun, Z.*, Preparation of disposable saliva α -amylase biosensor, *Chinese J. Anal. Chem.* **36** (2008) 1217. doi: [http://dx.doi.org/10.1016/S1872-2040\(08\)60068-7](http://dx.doi.org/10.1016/S1872-2040(08)60068-7)
24. *Shetty, V., Zigler, C., Robles, T. F., Elashoff, D., Yamaguchi, M.*, Developmental validation of a point-of-care, salivary α -amylase biosensor, *Psychoneuroendocrinology* **36** (2011) 193. doi: <http://dx.doi.org/10.1016/j.psyneuen.2010.07.008>
25. *Yamaguchi, M., Kanemaru, M., Kanemori, T., Mizuno, Y.*, Flow-injection-type biosensor system for salivary amylase activity, *Biosens. Bioelectron.* **18** (2003) 835. doi: [http://dx.doi.org/10.1016/S0956-5663\(03\)00007-1](http://dx.doi.org/10.1016/S0956-5663(03)00007-1)
26. *Gillard, B. K., Marksman, H. C., Feig, S. A.*, Direct spectrophotometric determination of alpha-amylase activity in saliva, with p-nitrophenyl alpha-maltoside as substrate, *Clin. Chem.* **23** (1977) 2279.
27. *Zajoncová, L., Jílek, M., Beranová, V., Pec, P.*, A biosensor for the determination of amylase activity, *Biosens. Bioelectron.* **20** (2004) 240. doi: <http://dx.doi.org/10.1016/j.bios.2004.01.006>
28. *Clem, D., Maidment, J., Ringham, J. M.*, A study into the measurement of alpha-amylase activity using phadebas, iodine and gel-diffusion procedures, *Nutr. Food Sci.* **31** (2001) 141. doi: <http://dx.doi.org/10.1108/00346650110385855>
29. *Gonzalez, C. F., Farina, J. I., Figueroa, L. I. C.*, A critical assessment of a viscometric assay for measuring *Saccharomycopsis fibuligera* alpha-amylase activity on gelatinised cassava starch, *Microb. Technol.* **30** (2002) 169. doi: [http://dx.doi.org/10.1016/S0141-0229\(01\)00479-3](http://dx.doi.org/10.1016/S0141-0229(01)00479-3)
30. *Sakač, N., Regušić, L., Sak-Bosnar, M., Jozanović, M., Breslauer, N.*, Direct potentiometric determination of ptyalin in saliva, *Int. J. Electrochem. Sci.* **9** (2014) 7097.